## Structure of Xanthomegnin and Related Pigments: Reinvestigation by <sup>13</sup>C Nuclear Magnetic Resonance Spectroscopy

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Summary The correct structures of xanthomegnin, rubrosulphin, and viopurpurin have been established as naphtho[2,3-c]pyran-8-yl dimers from <sup>13</sup>C n.m.r. data.

Xanthomegnin, first isolated from the mycelium of various *Trichophyton* spp., was assigned structure (1) from degradation studies and spectroscopic and chemical evidence.<sup>1</sup> Recently <sup>13</sup>C n.m.r. data were obtained for xanthomegnin produced by *Aspergillus melleus*<sup>2,3</sup> which seemed to provide additional support for the proposed structure.

According to our investigation of different naphtho- and anthra-quinones<sup>4</sup> the long-range coupling of the aromatic proton (e.g.,  $J_{\rm CH}$  5 Hz, to C-1) is in contradiction to structure (1). However, it is in good agreement with the isomeric structure (2) which also fits the chemical shift data since substituent-induced shifts are of the same order for ortho- and para-substituents. To prove the identity of the pigments obtained from different sources and the proposed structure (2) we have measured the proton-coupled and -decoupled <sup>13</sup>C n.m.r. spectra of authentic samples of xanthomegnin and its diacetate supplied by Professor G. Just.

The results given in the Table are identical with those reported by Simpson<sup>2</sup> thus confirming the existence of only one type of xanthomegnin with structure (2).† The minor

<sup>13</sup>C N.m.r. chemical shifts of xanthomegnin (2) and xanthomegnin diacetate (3) and <sup>13</sup>C-H coupling constants of (3) (δ in p.p.m. from external Me<sub>2</sub>Si,  $J_{CH}$  in Hz).<sup>a</sup>

Carbon	Compound (3)		
Carbon	δ	δ	$J_{ m CH}$
1	179.7	179.7	d, 4
<b>2</b>	157.9	$157 \cdot 7$	q, <b>3</b> ·5
3	122.7	124.0	s
4	$186 \cdot 1$	180.5	s
5	$162 \cdot 0$	150.9	S
6	$117 \cdot 2$	$125 \cdot 2$	S
7	$148 \cdot 2$	147.7	s
8	116.8	124.0	d, 168
9	134.6	135.8	s
10	114.5	123.0	d, 6
11	161.9	160.0	s
12	35.8	$34 \cdot 3$	t, <b>13</b> 0
13	74.4	$74 \cdot 1$	d, 152
14	20.4	19.6	q, 128
$2 ext{-OMe}$	$61 \cdot 2$	$60 \cdot 2$	q, 147
5-Ac		∫ 168· <b>3</b>	q, 3·5
		20.2	q, 1 <b>3</b> 0

<sup>a</sup> Bruker WH 270 spectrometer operating at 67.88 MHz was used; the spectrum of (2) was measured in (CD<sub>3</sub>)<sub>2</sub>SO—CDCl<sub>3</sub> (1:4) at 60 °C and that of (3) in (CD<sub>3</sub>)<sub>2</sub>SO at 100 °C. At 30 °C broad or doubled signals are observed as in the <sup>1</sup>H n.m.r. spectra (ref. 3).

<sup>†</sup> An independent proof of structure (2) could be obtained by oxidative dimerisation of semi-vioxanthin as originally suggested by us (A. Zeeck, personal communication).

differences observed for the corresponding acetate (± 1 p.p.m.) may be caused by the different conditions of the measurements.‡ As expected for (3), the C-1 and C-10 signals show long-range coupling to the proton at C-8  $(^3J_{\rm CH}$  4 and 6 Hz, respectively) while the acetoxy-bearing C-5, now separated by four bonds from this proton, gives a singlet. Broad signals are observed for C-6, C-7, and C-8 owing to multiple long-range coupling with the protons at C-12 and C-13.

Catalytic hydrogenation of (3) to the quinol was accompanied by rapid migration of the acetyl group to form (4) which is stabilized by a strong hydrogen bond to the lactonic carbonyl group. In the original work on xanthomegnin and related compounds1,3 the lowering of the lactonic carbonyl frequency on reduction of the corresponding acetates was thought to be a characteristic feature of the angular orientation of the  $\delta$ -lactone ring.

Since the related pigments viomellein,3 rubrosulphin,3 and viopurpurin<sup>3,5</sup> show the same <sup>13</sup>C chemical shifts as the corresponding xanthomegnin derivative for the naphthoquinone part as well as the characteristic long-range coupling of the aromatic proton, their structures have to be changed in the same way. As a consequence the biosynthesis of all members of the xanthomegnin family § can now be derived from a single intermediate, the dimeric naphthalene vioxanthin.5

We thank Professor G. Just for a generous gift of xanthomegnin derivatives and for references.

(Received, 17th April 1978; Com. 401.)

‡ The correct signal of C-6 (125·2 p.p.m.; in ref. 2, 132·7 p.p.m.) might have been overlooked because it is very broad at 30 °C.

§ This includes floccosin whose structure has been determined recently by X-ray analysis (F. Brisse, G. Just, and F. Blank, Acta Cryst., 1978, B34, 557).

<sup>1</sup>G. Just, W. C. Day, and F. Blank, Canad. J. Chem., 1963, 41, 74; A. S. Ng, G. Just, and F. Blank, ibid., 1969, 47, 1223.

<sup>2</sup> T. J. Simpson, *J.C.S. Perkin I*, 1977, 592.
<sup>3</sup> R. C. Durley, J. MacMillan, T. J. Simpson, A. T. Glen, and W. B. Turner, *J.C.S. Perkin I*, 1975, 163.

<sup>4</sup> G. Höfle, Tetrahedron, 1977, 33, 1963.

<sup>5</sup> F. Blank, A. S. Ng, and G. Just, Canad. J. Chem., 1966, 44, 2873.